RESEARCH ARTICLE

Open Access

¹⁸F-FDG uptake in the stomach on screening PET/CT: value for predicting *Helicobacter pylori* infection and chronic atrophic gastritis



Shigeki Kobayashi^{1*}, Mayumi Ogura¹, Naohisa Suzawa², Noriyuki Horiki³, Masaki Katsurahara³, Toru Ogura⁴ and Haiime Sakuma²

Abstract

Background: The aim of this study was to determine the value of ¹⁸F-FDG uptake on screening PET/CT images for the prediction of *Helicobacter pylori* (*H. pylori*) infection and chronic atrophic gastritis.

Methods: Among subjects who underwent ¹⁸F-FDG PET/CT for cancer screening from April 2005 to November 2015, PET/CT images were analyzed in 88 subjects who had gastrointestinal fiberscopy within 6 months. The volumes of interest (VOIs) were placed in the fornix, corpus and antrum of the stomach to determine maximal standardized uptake value (SUVmax) and mean SUV (SUVmean). Receiver operating characteristic curve (ROC) analysis was performed to determine the diagnostic performance of SUV indicators in predicting *H. pylori* infection and chronic atrophic gastritis.

Results: SUV indicators of the stomach were significantly higher in subjects with H. pylori infection than those without (from P < 0.001 to P < 0.05). ROC analysis revealed that SUVmean had the highest performance in predicting H. pylori infection (AUC 0.807) and chronic atrophic gastritis (AUC 0.784). SUVmean exhibited the sensitivity of 86.5 % and the specificity of 70.6 % in predicting H. pylori infection, and the sensitivity of 75.0 % and 78.6 % in predicting chronic atrophic gastritis.

Conclusion: Assessment of ¹⁸F-FDG uptake in the stomach reflecting active inflammation is useful in predicting patients with *H. pylori* infection and subsequent chronic atrophic gastritis which is closely associated with the risk of gastric neoplasms.

Keywords: ¹⁸F-FDG PET/CT, Helicobacter pylori infection, Chronic atrophic gastritis

Background

Helicobacter pylori (H. pylori) infection is strongly related with many gastroduodenal diseases including peptic ulcer diseases, chronic atrophic gastritis, mucosa associated lymphoid tissue (MALT) lymphoma and gastric cancer [1, 2]. In particular, gastric cancer is the third most common of all cancers among males and the fifth most common among females. Once infection of H. pylori is established, it usually lasts for life and

exhibits carcinogenicity which induces gastric cancer through chronic atrophic gastritis [3].

¹⁸F-FDG PET/CT is widely used in cancer staging and cancer screening. However, previous studies demonstrated that the sensitivity of ¹⁸F-FDG-PET in screening gastric cancer in asymptomatic subjects was limited, ranging from 10 % to 38 % [4, 5]. The main difficulty in ¹⁸F-FDG-PET diagnosis of gastric cancer is attributed to physiological uptake of ¹⁸F-FDG in the stomach [6–10]. In addition to the abnormal ¹⁸F-FDG uptake associated to malignant tumors, physiological or inflammation related uptakes are seen on ¹⁸F-FDG PET images. Takahashi et al [11] evaluated the pattern of ¹⁸F-FDG uptake in

Full list of author information is available at the end of the article



^{*} Correspondence: skhousya@clin.medic.mie-u.ac.jp

¹Center for Preventive Medicine, Mie University Hospital, 2-174 Edobashi, Tsu, Mie Pref. 514-8507, Japan

the stomach in association with endoscopic findings of the gastric mucosa in 272 cases and found that accumulation pattern of ¹⁸F-FDG largely corresponds to the presence of mucosal inflammation. Although semiquantitative evaluation of ¹⁸F-FDG uptake using standardized uptake values (SUVs) in the stomach has been used for assessing MALT lymphoma [12], differentiating malignant and benign gastric diseases [13] and predicting the prognosis of gastric carcinoma [14], the value of SUV measurement of FDG uptake for detecting H. pylori infection and subsequent chronic atrophic gastritis has not been well established. Lin et al [15] found a significant positive correlation between SUVs of ¹⁸F-FDG in the stomach and the values of C-13 urea breath test which is the most commonly used noninvasive test for H. pylori. However, the number of the subjects was limited (n = 16) and endoscopic examination was not performed in their study.

Consequently, the aim of this study was to investigate the value of semi-quantitative assessment of ¹⁸FDG uptake in the stomach with SUV for predicting *H. pylori* infection and chronic gastritis in subjects who underwent ¹⁸F-FDG PET/CT for cancer screening.

Methods

Subjects

Medical records of subjects who underwent ¹⁸F-FDG PET/CT for cancer screening between April 2005 and November 2015 were retrospectively investigated. Among them, 88 subjects underwent gastrointestinal fiberscopy within 6 months of the PET/CT study. The reasons for gastrointestinal fiberscopy were increased uptake of ¹⁸F-FDG in the stomach (55 subjects), previous history of peptic ulcer (5 subjects), familial history of peptic ulcer or gastric cancer (4 subjects), or request by the examinee (32 subjects). None of these 88 subjects had a previous history of gastric cancer or MALT lymphoma. The presence or absence of *H. pylori* infection, as well as the diagnosis of chronic atrophic gastritis, were determined in the medical records in all 88 subjects who underwent gastrointestinal fiberscopy. Thus, ¹⁸F-FDG PET/CT images were evaluated in these 88 subjects. This retrospective study was approved by the institutional review board of Mie University Hospital (study no. 2989) and was conducted in accordance with the Declaration of Helsinki and Good Clinical Practice. Informed consent was waived for this retrospective study.

PET/CT imaging

All subjects fasted for at least 6 h before PET/CT acquisitions. Prior to ¹⁸F-FDG injection, blood glucose levels were determined from capillary blood samples and were confirmed to be less than 150 mg/dl in all subjects. A

3.7-MBq/kg dose of ¹⁸F-FDG was injected intravenously in one arm. PET/CT was performed by using an Aquiduo PCA-700B scanner (Toshiba, Nasu, Japan) or Discovery PET/CT 690 scanner (GE, Milwaukee, WI). Images from the skull to the mid-thigh were acquired approximately 60 min after ¹⁸F-FDG injection, by employing 3-dimentional acquisitions in 7-9 bed positions with 2-min acquisition in each position. Subjects were placed supine with the arms alongside the body or lifted up to the skull and were allowed to breathe normally during PET acquisitions. CT images acquired in approximately ten seconds during a natural breathholding were used for attenuation correlation and generation of fusion images. Attenuation-corrected PET images with co-registered CT data were reviewed.

PET/CT image analysis

¹⁸F-FDG uptake in the stomach was measured semiquantitatively by placing volumes of interest (VOIs) at the fornix, corpus and antrum of the stomach as well as in the liver by consensus of two observers. The VOIs were 3D spheres and the size of VOIs were 5 mm in diameter for the stomach, and 30 mm in diameter for the liver. The VOI for the stomach was carefully placed in the gastric wall by monitoring both PET-CT fusion images and PET images. VOI for the liver was placed to avoid the region just blow the diaphragm for preventing the motion blurring artifact. For each VOI, maximal SUV (SUVmax) and mean SUV (SUVmean) were recorded (Fig. 1). ROC analysis was performed for several different SUV indicators. Maximum SUVmax and mean SUVmax were the maximum and mean values of the SUVmax measured at the fornix, corpus and antrum. Maximum SUVmean and mean SUVmean were the maximum and mean values of the SUVmean measured at the fornix, corpus and antrum. In addition, maximum SUVmax / SUVmean liver, mean SUVmax /SUVmean liver, maximum SUVmean / SUVmean liver and mean SUVmean / SUVmean liver were determined to evaluate the diagnostic performance of these indicators in predicting *H pylori* infection and chronic atrophic gastritis.

Statistical analysis

SPSS version 22.0 software (IBM Japan, Tokyo, Japan) was used for statistical analyses. We determined whether statistically significance difference was observed in SUVs of the stomach between those with and without *H. pylori*, and those with and without chronic atrophic gastritis. The sensitivity and specificity of SUV indicators in predicting *H. pylori* infection and chronic atrophic gastritis were calculated by using an optimal cut-off point on the ROC curve that has the minimum distance to the upper left corner (where sensitivity = 1 and specificity = 1). The statistically significance was evaluated by Mann-

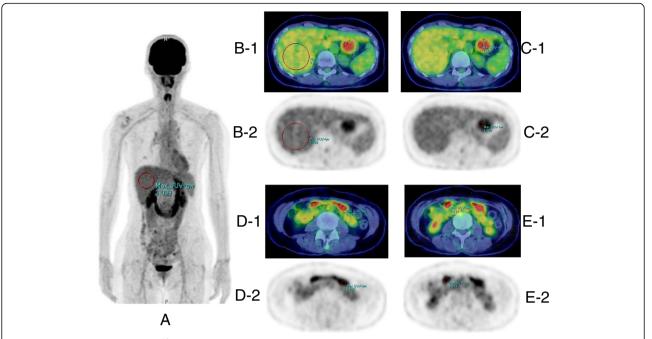


Fig. 1 The methods for measuring ¹⁸F-FDG uptake of the stomach and the liver. VOIs of 3D sphere were placed at the fornix, corpus and antrum of the stomach and the liver in each subject. SUVmax (shown as Max SUV-bw on MIP, fusion images and PET images) and SUVmean were determined in each VOI in the stomach, and SUVmean liver was determined in liver VOI. **a** A MIP image of a subject with *H. pylori* infection, VOI was placed to avoid the area just blow the diaphragm for preventing the motion blurring artifact. The VOIs were placed by monitoring both PET/CT fusion images and PET images. **b-1**, 2 VOI of the liver. **c1**, 2 VOI of gastric fornix. **d-1**, 2 VOI of gastric corpus. **e-1**, 2 VOI of gastric antrum

Whitney *U*-test or Wilcoxon rank sum test. All analysis were 2-sided, a P-value of less than 0.05 was considered statistically significant.

Results

Characteristics of subjects

Characteristics of the subjects including laboratory diagnosis by gastrointestinal fiberscopy are shown in Table 1. Diagnosis of *H. pylori* infection was made by a rapid urease test, a stool antigen test and an information of previous medical institution or prevention center. Three subjects who had chronic atrophic gastritis without *H. pylori* infection on medical records in previous medical institutions, were turned out to be H. pylori positive by further investigation in our hospital.

PET/CT image analysis

Table 2 summarizes the SUVmax and SUVmean of 18 F-FDG uptake at the fornix, corpus and antrum, as well as the maximum and the mean values of SUVmax and SUVmean at 3 regions in the stomach in associated with $H.\ pylori$ infection. Table 3 summarizes these SUV indicators in associated with chronic atrophic gastritis. All of these SUV indicators in the stomach were significantly higher in patients with $H.\ pylori$ infection than in those without $H.\ pylori$ (P < 0.001). In addition, all of these SUV indicators were significantly higher in

patients with chronic atrophic gastritis than in those without chronic atrophic gastritis (P < 0.001). It was also noted that the 18 F-FDG uptake of the fornix in the stomach was significantly higher than those in corpus and antrum, independent of the presence of H. pylori infection and chronic atrophic gastritis.

Diagnostic performance by ROC analysis

Figure 2 shows ROC curves for SUV indicators in predicting *H. pylori* infection and chronic atrophic gastritis. In Table 4, the area under ROC curves, the optimal cutoff values, the sensitivities and specificities in predicting H. pylori infection and chronic atrophic gastritis are presented. All of SUV indicators demonstrated good diagnostic performance for the prediction of H. pylori infection and chronic atrophic gastritis. Among these 4 SUV indicators, mean SUVmean exhibited the highest area under ROC curves for predicting H. pylori infection (0.807, 95%CI 0.715 - 0.898) and for chronic gastritis (0.784, 95 % CI 0.684 – 0.884). As shown in Table 5, normalization of these SUV indicators in the stomach by the liver SUV did not improve the area under ROC curves for the diagnosis of H. pylori infection. For predicting chronic atrophic gastritis, normalization of SUV indicators by the liver SUV slightly improve the area under ROC curves, with the highest area under ROC curve of 0.793 (95 % CI 0.686 - 0.900) by mean

Table 1 Characteristics of the subjects

Age (y)	
Mean ± SD	58 ± 11
Range	34 – 79
Gender	Number (%)
Female	38 (43.2)
Male	50 (56.8)
H. Pylori infection	Number (%)
Positive	37 (42.0)
Negative	51 (58.0)
Chronic atrophic gastritis	Number (%)
Positive (H. Pylori positive)	37 (40.9)
(H. Pylori negative)	24 (27.3)
Negative	27 (31.8)
Neoplasms (finding on fiberscopy)	Number
Early gastric cancer (H. Pylori positive)	4
Gastric adenoma (H. Pylori positive)	2
MALT lymphoma (H. Pylori positive)	1
Other fiberscopic findings	3
Superficial gastritis (H. Pylori positive)	3
(H. Pylori negative)	2
Erosive gastritis (H. Pylori positive)	2
(H. Pylori negative)	3
Gastric ulcer scar (H. Pylori positive)	1
Duodenal ulcer scar (H. Pylori positive)	1
Erosion of E-C Junction (H. Pylori negative)	1
Reflux esophagitis (H. Pylori positive)	1
(H. Pylori negative)	1

Table 2 The SUVs of 18 F-FDG in the stomach in associated with *Helicobacter pylori* infection

With Thenedoucter pyron infection					
	H. pylori(+) N = 37	H. pylori(-) N = 5			
SUVmax (mean ± S	D)				
Fornix	4.01 ± 0.80	$3.38 \pm 0.97^{++}$			
Corpus	$3.70 \pm 0.95^{**}$	$2.71 \pm 0.90^{*+}$			
Antrum	$3.58 \pm 1.12^{***}$	$2.68 \pm 0.99^{*+}$			
Maximum	4.36 ± 0.88	3.57 ± 1.01			
Mean	3.76 ± 0.78	$2.93 \pm 0.80^{+}$			
SUVmean (mean ±	SD)				
Fornix	3.62 ± 0.71	$3.06 \pm 0.90^{++}$			
Corpus	$3.31 \pm 0.87^{***}$	$2.39 \pm 0.83^{*+}$			
Antrum	$3.06 \pm 0.96^{**}$	$2.30 \pm 0.87^{*+}$			
Maximum	3.90 ± 0.74	$3.19 \pm 0.92^{+}$			
Mean	3.33 ± 0.67	$2.58 \pm 0.73^{+}$			

Significant difference between each region and fornix at same group (*P < 0.001, **P < 0.01, ***P < 0.05)

Significant difference between H. Pylori (+) and H. Pylori (-) at same region (+P < 0.001, +++P < 0.01, +++P < 0.05)

Table 3 The SUVs of 18 F-FDG in the stomach in associated with chronic gastritis

9		
	Chr Gastritis(+) $N = 60$	Chr Gastritis(-) N = 28
SUVmax (mean ±	± SD)	
Fornix	3.86 ± 0.91	$3.20 \pm 0.92^{+}$
Corpus	$3.39 \pm 1.00^*$	$2.57 \pm 0.92^{*+}$
Antrum	$3.37 \pm 1.19^{**}$	$2.39 \pm 0.67^{*+}$
Maximum	4.12 ± 0.97	$3.42 \pm 1.02^{++}$
Mean	3.54 ± 0.89	$2.72 \pm 0.62^{+}$
SUVmean (mean	± SD)	
Fornix	3.49 ± 0.85	$2.88 \pm 0.80^{+++}$
Corpus	$3.02 \pm 0.94^*$	$2.25 \pm 0.81^{*+}$
Antrum	$2.88 \pm 1.01^*$	$2.06 \pm 0.65^{*+}$
Maximum	3.69 ± 0.88	$3.06 \pm 0.90^{++}$
Mean	3.13 ± 0.79	$2.40 \pm 0.55^{+}$

Significant difference between each region and fornix at same group (*P < 0.001, **P < 0.01)

Significant difference between Chr Gastritis (+) and Chr Gastritis (-) at same region (+ P < 0.001, ++ P < 0.01, +++ P < 0.05)

SUVmax. However, the amount of improvement with normalization by the liver SUV was quite limited.

Dot plots of mean SUVmean in subjects with and without *H. pylori* infection and in those with and without chronic gastritis were shown on Fig. 3. The sensitivity and specificity of mean SUVmean were 86.5 % and 70.6 % for *H. pylori* infection (optimal cut-off value of 2.66), and 75.0 % and 78.6 % for chronic gastritis (optimal cut-off value of 2.57), respectively.

Gastric neoplasms found by GIF

Among the 88 subjects, seven neoplasms were found on gastrointestinal fiberscopy, including four early gastric cancers, two gastric adenomas and a MALT lymphoma. In four gastric cancer and two gastric adenomas, no focal increase in ¹⁸F-FDG uptake corresponding to tumors was observed, while *H. pylori* was positive in these cases. In a patient with MALT lymphoma, the antibody test of *H. pylori* was negative and increased focal ¹⁸F-FDG uptake at gastric corpus was detected which corresponded to MALT lymphoma proven by gastrointestinal fiberscopy. The infection of *H. pylori* was also demonstrated by histologic specimen taken by fiberscopy.

Discussion

In the current study, we investigated the value of 18 F-FDG uptake measured by SUV on screening PET/CT images for the prediction of *H. pylori* infection and chronic atrophic gastritis determined by gastrointestinal fiberscopy. The major findings in this study were [1] The SUV of 18 F-FDG uptake in the stomach was significantly elevated in patients with *H. pylori* infection and in those with chronic atrophic gastritis [2]; 18 F-

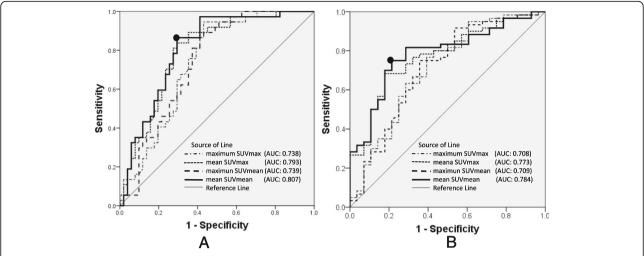


Fig. 2 ROC curves for SUV indicators. **a** ROC curves for predicting *H. pylori* infection. **b** ROC curves for predicting chronic atrophic gastritis. Among these SUV indicators, the highest diagnostic performance was achieved with the mean SUVmean in the fornix, corpus and antrum for predicting *H. pylori* infection as well as for predicting chronic atrophic gastritis

FDG uptake of the fornix in the stomach was significantly higher than those in corpus and antrum regardless of *H. pylori* infection and chronic atrophic gastritis [3]; mean SUVmean showed the highest area under ROC curves for predicting *H. pylori* infection (0.807) and chronic atrophic gastritis (0.784), and is useful for identifying patients who require gastrointestinal fiberscopy. Normalization of stomach SUVs by liver SUV provided minimal differences in the diagnostic performance and is not considered to be necessary.

Accumulation of ¹⁸F-FDG in the stomach

Pattern of accumulation of ¹⁸F-FDG in the stomach and its associated with endoscopic findings of the gastric mucosa and *H. pylori* infection were previously investigated by Takahashi et al [11] by using a visual assessment of ¹⁸F-FDG PET image. They classified ¹⁸F-FDG uptake in the stomach into three groups (A: localized accumulation in the fornix, B: diffuse accumulation throughout the entire stomach, *C*: no accumulation).

They found that *H. pylori* infections were more frequent in Groups A and B than in Group C, concluding that accumulation of ¹⁸F-FDG in the stomach suggests a high probability of inflammatory changes to the gastric mucosa, forming a background for the development of cancer or malignant lymphoma. In our current study, we used a more objective approach by measuring SUVs of ¹⁸F-FDG in the fornix, corpus and antrum. Consistent with previous report [8, 11], we found that ¹⁸F-FDG uptake of the fornix was significantly higher than corpus and antrum. In addition, SUV of ¹⁸F-FDG in the fornix was significantly higher than those in corpus and antrum, not only in the subjects with H. pylori infection and chronic atrophic gastritis, but also in those without H. pylori infection or chronic atrophic gastritis, suggesting that high ¹⁸F-FDG uptake in an oral side of the stomach is physiological. We also noticed that H. pylori infection and chronic atrophic gastritis are associated with elevated SUVs in all gastric regions including the fornix, corpus and antrum. This indicates that H. pylori

Table 4 Diagnostic performance of SUVs for *H. pylori* infection

Predictive Indicators	AUC	Cut-off	Sensitivity	Specificity	95 % CI of AUC	P value
Maximun SUVmax	0.738	3.66	81.1 %	60.8 %	0.635 - 0.841	<0.001
Mean SUVmax	0.793	3.11	81.1 %	72.5 %	0.699 – 0.887	< 0.001
Maximun SUVmean	0.739	3.30	81.1 %	62.7 %	0.636 - 0.841	< 0.001
Mean SUVmean	0.807	2.66	86.5 %	70.6 %	0.715 - 0.898	< 0.001
Diagnostic performance of	SUVs for chronic a	trophic gastritis				
Maximun SUVmax	0.708	3.42	76.7 %	60.7 %	0.585 - 0.831	0.02
Mean SUVmax	0.773	2.86	76.7 %	67.9 %	0.671 - 0.875	< 0.001
Maximun SUVmean	0.709	3.15	75.0 %	64.3 %	0.585 - 0.833	0.02
Mean SUVmean	0.784	2.57	75.0 %	78.6 %	0.684 - 0.884	< 0.001

Table 5 Brightosite performance of 50 to Hormanized by 50 to Hit the liver for 11. Tylon infection						
Predictive Indicators	AUC	Cut-off	Sensitivity	Specificity	95 % CI of AUC	P value
Maximun SUVmax	0.739	1.64	64.9 %	72.5 %	0.637 - 0.841	< 0.001
Mean SUVmax	0.796	1.31	81.1 %	74.5 %	0.700 - 0.892	< 0.001
Maximum SUVmean	0.738	1.37	83.8 %	60.8 %	0.635 - 0.841	< 0.001
Mean SUVmean	0.791	1.15	81.1 %	72.5 %	0.695 - 0.887	< 0.001
Diagnostic performance of S	SUVs normalized b	y SUV in the liver	for chronic atrophic ga	astritis		
Maximun SUVmax	0.721	1.44	73.3 %	60.7 %	0.596 – 0.847	0.01
Mean SUVmax	0.793	1.25	76.7 %	78.6 %	0.686 - 0.900	< 0.001
Maximum SUVmean	0.711	1.25	83.3 %	60.7 %	0.583 - 0.838	0.02
Mean SUVmean	0.790	1.09	78.3 %	75.0 %	0.682 - 0.897	< 0.001

Table 5 Diagnostic performance of SUVs normalized by SUV in the liver for H. Pylori infection

infection causes increased $^{18}\text{F-FDG}$ uptake reflecting active inflammation throughout the entire stomach.

Detection of gastric neoplasms by ¹⁸F-FDG PET/CT

For the assessment of patients with advanced gastric cancer, ¹⁸F-FDG PET/CT has been shown to be useful in detecting nodal metastasis and distant metastasis, and in predicting prognosis [16–20]. However, ¹⁸F-FDG PET/CT is not useful for screening gastric cancers [5, 11, 21, 22]. Shoda et al. studies 2861 asymptomatic subjects and found that the sensitivity of ¹⁸F-FDG PET for gastric cancer was as low as 10 % [3]. Consequently the use of gastrointestinal fiberscopy is considered more appropriate in screening gastric cancer.

Clinical implications

Our results demonstrated that semi-quantitative assessment of ¹⁸F-FDG uptake with SUV has high diagnostic accuracy in predicting *H. pylori* infection and chronic atrophic gastritis. As previously mentioned, *H. pylori* infection and subsequent chronic atrophic gastritis lead to increased risk of gastric cancer formation. Inflammatory

change in the gastric mucosa caused by H. pylori forms a background for the development of gastric cancer or malignant lymphoma. In a Japanese cohort study, the population attributable fraction (PAF) of H. pylori infection for gastric cancer incidence (i.e. the fraction of gastric cancer incident cases that is attributable to *H. pylori* infection) was estimated to be 84 % [3]. Despite the declined prevalence of *H. pylori* infection for the past 30 years, gastric cancer is the second most frequent cause of cancer death in both males and females in Japan, and the most frequent cancer in males and the second most frequent cancer in females [23]. Therefore, gastrointestinal fiberscopy should be strongly recommended for subjects with increased ¹⁸F-FDG uptake in the stomach. According to the results in this study, high area under ROC of 0.807 and high sensitivity of 86.5 % can be achieved when mean SUVmean values of > 2.66 was used as a threshold.

Limitations

Several limitations must be acknowledged in this study. First, this is a single-center study with a limited number

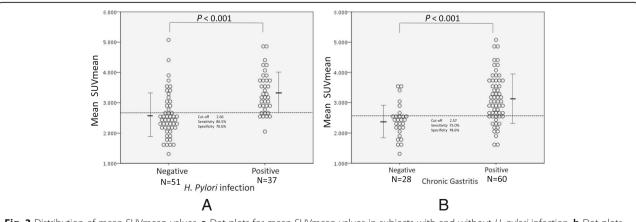


Fig. 3 Distribution of mean SUVmean values. **a** Dot plots for mean SUVmean values in subjects with and without H. pylori infection. **b** Dot plots for mean SUVmean values in subjects with and without chronic atrophic gastritis. Statistical significant difference for the mean SUVmean values was observed between subjects with and without H. pylori infection (p < 0.001) and between subjects with and without chronic qastritis (p < 0.001)

of subjects, and there is a selection bias for subjects who underwent gastrointestinal fiberscopy. Second, the degree of chronic atrophic gastritis was not evaluated in current study, because the laboratory diagnosis by gastrointestinal fiberscopy was qualitative and operator-dependent. Third, CT images were acquired during natural breath-holding while PET images were obtained during free-breathing. This may result in misregistration artifact and alteration in SUV. Forth, SUV of ¹⁸F-FDG uptake in the stomach was not compared with the gastrointestinal fiberscopy findings in detail. Further investigation by prospective multi-center study using both PET-CT and gastrointestinal fiberscopy is necessary to determine the value of ¹⁸F-FDG PET in early detection and prevention of gastric cancer.

Conclusion

Uptake of ¹⁸F-FDG in the stomach reflecting active inflammation is strongly associated with *H. pylori* infection and subsequent chronic atrophic gastritis. Subjects demonstrating increased SUV of ¹⁸F-FDG uptake in the stomach should be recognized as patients with high likelihood *H. pylori* infection and at increased risk of gastric neoplasms. Gastrointestinal fiberscopy should be recommended in these subjects.

Abbreviations

¹⁸F-FDG PET/CT: [¹⁸F] fluoro-2-deoxy-D-glucose positron emission tomography/computed tomography; AUC: Area under the curve; CI: Confidence interval; FDG: Fluoro-2-deoxy-D-glucose; MALT: Mucosa associated lymphoid tissue; PET/CT: Positron emission tomography/computed tomography; ROC: Receiver operating characteristic curve; SUV: Standardized uptake value; SUVmax: Maximal standardized uptake value; SUVmean: Mean standardized uptake value

Acknowledgements

We would like to thank the whole Nuclear Medicine imaging technologist team at Mie university Hospital, for continuous support. We are also would like thank the center for Preventive Medicine team at Mie University Hospital for continuous cooperation. All the authors were supported by the Mie University Hospital in the study.

Funding

None.

Availability of data and materials

Data to replicate findings are in the Figures and Tables of the main paper. Due to patient privacy protection, any additional materials of the study are only available upon individual request directed to the corresponding author.

Author's contributions

SK organized the entire study, participated in the study design, evaluated the results and wrote the first and revised manuscript. MO participated in the study design and directed PET/CT examinations. NS carried out the images analysis with SK for the first and revised manuscripts. NH and MK participated in the study design, and directed and carried out gastrointestinal fiberscopy. TO performed the data analysis and statistical analysis in the first and revised manuscripts. HS participated in the design of the study, redesigned the data analysis in the revision and edited the first and revised manuscripts. All authors read and approved the final manuscript.

Competing interests

The authors declare that they have no competing of interest.

Consent for publication

Not applicable.

Ethics approval and consent to participate

Current study was approved by the Institutional Review Board (IRB) at the corresponding author's institution (study no. 2989) and need for signed informed consent was waived.

Author details

¹Center for Preventive Medicine, Mie University Hospital, 2-174 Edobashi, Tsu, Mie Pref. 514-8507, Japan. ²Department of Radiology, Mie University Hospital, 2-174 Edobashi, Tsu, Mie Pref. 514-8507, Japan. ³Department of Gastroenterology and Hepatology, Mie University Hospital, 2-174 Edobashi, Tsu, Mie Pref. 514-8507, Japan. ⁴Clinical Research Support Center, Mie University Hospital, 2-174 Edobashi, Tsu, Mie Pref. 514-8507, Japan.

Received: 4 February 2016 Accepted: 13 October 2016 Published online: 18 October 2016

References

- Tally NJ, Zinsmeister AR, Weaver A, et al. Gastric adenocarcinoma and Helicobacter pylori infection. J Natl Cancer Inst. 1991;83:1734–9.
- Sokic-Milutinovic A, Alempijevic T, Milosavljevic T. Role of Helicobacter pylori infection on gastric carcinogenesis: Current knowledge and future directions. World J Gastroenterol. 2015;21:11654

 –72.
- Watanabe M, Ito H, Hosono S, et al. Declining trends in prevalence of Helicobacter pylori infection by birth-year in a Japanese population. Cancer Sci. 2015;16(12):1738–43.
- Shoda H, Kakugawa Y, Saito D, et al. Evaluation of ¹⁸F-2-deoxy-2-fluoro-glucose positron emission tomography for gastric cancer screening in asymptomatic individuals undergoing endoscopy. Br J Cancer. 2007;97:1493–8.
- Minamimoto R, Senda M, Jinnouchi S, et al. Performance profile of a FDG-PET cancer screening program for detecting gastric cancer: results from a nationwide Japanese survey. Jpn J Radiol. 2014;32:253–9.
- Shreve PD, Anzai Y, Wahl RL. Pitfalls in oncologic diagnosis with FDG PET imaging: physiologic and benign variants. RadioGraphics. 1999;19:61–77.
- Zhuang H, Alavi A. 18-Fluorodeoxyglucose positron emission tomographic imaging in the detection and monitering of infection and inflammation. Semin Nucl Med. 2002;17:47–59.
- Koga H, Sasaki M, Kuwabara Y, et al. An analysis of the physiological FDG uptake pattern in the stomach. Ann Nucl Med. 2003;17:733–8.
- Kamel EM, Thumshirn M, Truninger K, et al. Significance of incidental 18 F-FDG accumulations in the gastrointestinal tract in PET/CT: correlation with endoscopic and histopathologic results. J Nucl Med. 2004;45:1804–10.
- Israel O, Yefremov N, Bar-Shalom R, et al. PET/CT detection of unexpected gastrointestinal foci of 18 F-FDG uptake: incidence, localization patterns, and clinical significance. J Nucl Med. 2005;46:758–62.
- Takahashi H, Ukawa K, Ohkawa N, et al. Significance of ¹⁸F-2-deoxy-2-fluoroglucose accumulation in the stomach on positron emission tomography. Ann Nucl Med. 2009;23:391–7.
- Hirose Y, Kaida H, Ishibashi M, et al. Comparison between endoscopic macroscopic classification and F-18 FDG PET findings in gastric mucosaassociated lymphoid tissue lymphoma patients. Clin Nucl Med. 2012;37:152–7.
- Cui J, Zhao P, Ren MZ, et al. Evaluation of dual time point imaging ¹⁸F-FDG PET/CT in differentiating malignancy from benign gastric diseases. Medicine. 2015;94:1–5.
- Grabinska K, Pelak M, Wydmanski J, et al. Prognostic value and clinical correlations of 18-F-fluorodeoxyglucose metabolism quantifiers in gastric cancer. World J Gastroenterol. 2015;21:5901–9.
- Lin CY, Liu CS, Ding HJ, et al. Positive correlation between standardized uptake values of FDG uptake in the stomach and the value of the C-13 urea test. Clin Nucl Med. 2006;31:792–4.
- Kim HW, Won KS, Song BI, et al. Correlation of primary tumor FDG uptake with histopathologic features of advanced gastric cancer. Nucl Med Mol Imaging. 2015;49:135–42.
- Song BI, Kim HW, Won KS, et al. Preoperative standardized uptake value of metastatic lymph nodes measured by ¹⁸F-FDG PET/CT improves the prediction of prognosis in gastric cancer. Medicine. 2015;94:1–8.
- Altini C, Asabella NA, Palo AD, et al. ¹⁸F-FDG PET/CT role in staging of gastric carcinomas: Comparison with conventional contrast enhancement computed tomography. Medicine. 2015;94:1–8.

- Filik M, Kir KM, Aksel B, et al. The role of ¹⁸F-FDG PET/CT in the primary staging of gastric cancer. Mol Imaging Radionuclide Therapy. 2015;24:15–20.
- Yun M. Imaging of gastric cancer metabolism using ¹⁸F-FDG PET/CT. J Gastric Cancer. 2014;14:1–6.
- 21. Wang YK, Kuo FC, Liu CJ, et al. Diagnosis of Helicobacter pylori infection: Current options and developments. World J Gastroenterol. 2015;21:11221–35.
- 22. Malibari N, Hickeson M, Lisbona R. PET/computed tomography in the diagnosis and staging gastric cancers. PET Clin. 2015;10:311–26.
- 23. The editorial board of the cancer statistics in Japan. Cancer statistics in Japan 2014. Foundation for Promotion of Cancer Research. 2014:1-115

Submit your next manuscript to BioMed Central and we will help you at every step:

- We accept pre-submission inquiries
- Our selector tool helps you to find the most relevant journal
- We provide round the clock customer support
- Convenient online submission
- Thorough peer review
- Inclusion in PubMed and all major indexing services
- Maximum visibility for your research

Submit your manuscript at www.biomedcentral.com/submit

