RESEARCH

Open Access

Quantitative muscle MRI displays clinically relevant myostructural abnormalities in long-term ICU-survivors: a case–control study

R. Rehmann^{1*}, E. Enax-Krumova¹, C. H. Meyer-Frießem^{2†} and L. Schlaffke^{1†}

Abstract

Background Long-term data on ICU-survivors reveal persisting sequalae and a reduced quality-of-life even after years. Major complaints are neuromuscular dysfunction due to Intensive care unit acquired weakness (ICUAW). Quantitative MRI (qMRI) protocols can quantify muscle alterations in contrast to standard qualitative MRI-protocols.

Methods Using qMRI, the aim of this study was to analyse persisting myostructural abnormalities in former ICU patients compared to controls and relate them to clinical assessments. The study was conducted as a cohort/ case–control study. Nine former ICU-patients and matched controls were recruited (7 males; 54.8y \pm 16.9; controls: 54.3y \pm 11.1). MRI scans were performed on a 3T-MRI including a mDTI, T2 mapping and a mDixonquant sequence. Water T2 times, fat-fraction and mean values of the eigenvalue (λ_1), mean diffusivity (MD), radial diffusivity (RD) and fractional anisotropy (FA) were obtained for six thigh and seven calf muscles bilaterally. Clinical assessment included strength testing, electrophysiologic studies and a questionnaire on quality-of-life (QoL). Study groups were compared using a multivariate general linear model. qMRI parameters were correlated to clinical assessments and QoL questionnaire using Pearson's correlation.

Results qMRI parameters were significantly higher in the patients for fat-fraction (p < 0.001), water T2 time (p < 0.001), FA (p = 0.047), MD (p < 0.001) and RD (p < 0.001). Thighs and calves showed a different pattern with significantly higher water T2 times only in the calves. Correlation analysis showed a significant negative correlation of muscle strength (MRC sum score) with FA and T2-time. The results were related to impairment seen in QoL-questionnaires, clinical testing and electrophysiologic studies.

Conclusion qMRI parameters show chronic next to active muscle degeneration in ICU survivors even years after ICU therapy with ongoing clinical relevance. Therefore, qMRI opens new doors to characterize and monitor muscle changes of patients with ICUAW. Further, better understanding on the underlying mechanisms of the persisting complaints could contribute the development of personalized rehabilitation programs.

Keywords Critical illness neuropathy, Critical illness myopathy, ICUAW, Quantitative MRI, T2 mapping, Muscle diffusion tensor imaging, Fat fraction

[†]C. H. Meyer-Frießem and L. Schlaffke both authors contributed equally.

*Correspondence: R. Rehmann Robert.Rehmann@rub.de Full list of author information is available at the end of the article



© The Author(s) 2023. **Open Access** This article is licensed under a Creative Commons Attribution 4.0 International License, which permits use, sharing, adaptation, distribution and reproduction in any medium or format, as long as you give appropriate credit to the original author(s) and the source, provide a link to the Creative Commons licence, and indicate if changes were made. The images or other third party material in this article are included in the article's Creative Commons licence, unless indicated otherwise in a credit line to the material. If material is not included in the article's Creative Commons licence and your intended use is not permitted by statutory regulation or exceeds the permitted use, you will need to obtain permission directly from the copyright holder. To view a copy of this licence, visit http://creativecommons.org/licenses/by/A.0/. The Creative Commons Public Domain Dedication waiver (http://creativecommons.org/publicdomain/zero/1.0/) applies to the data made available in this article, unless otherwise stated in a credit line to the data.

Background

Intensive care unit acquired weakness (ICUAW) is a general term that integrates the clinical and pathophysiological aspects of symmetric distal axonal neuropathy (CIP) and proximal myopathy (CIM) related to an intensive care therapy. ICUAW is common in intensive care patients (up to 82% of patients) and is an independent risk factor for long-term disability and a reduced quality of life in ICU survivors [1-3]. CIM is thought to be the result of muscle protein breakdown and myofiber necrosis due to inflammatory systemic responses and inactivity [4-6]. In contrast to CIP, CIM is usually transient and resolves during rehabilitation. In CIP systemic toxins, inflammatory responses, endothelial changes in sepsis as well as vasoactive and other medication are prone to cause microvascular damage and toxic axonal degeneration that is often non-reversible and cause long-lasting clinical deficits [7]. Thus, ICUAW can be regarded as an organ failure of the peripheral neuromuscular system with a high impact on long-term disability [1, 2].

Long-term data of ICU survivors are rare. A 5-year follow-up revealed a reduced motor performance and quality of life [3, 8–10]. Thus, objective outcome measures are highly important to quantify the disease status of the patients. For the measurement of disability, especially in CIP—since it contributes mainly to long-lasting dysfunction—MRC grading, walking tests, electrophysiologic testing, quality of life questionnaires and patients functional status have been used [1, 2, 10–13].

Van Aerde et al. showed that even a slight reduction in MRC sum score could be associated with a high 5-year morbidity and mortality after ICU discharge and underline the importance to capture muscle degeneration early [14]. Imaging studies in ICUAW mostly relate to computed tomography (CT) and muscle ultrasound (US) where a reduction in muscle mass and a change in muscle composition can reflect muscle degeneration due to ICUAW [15–19]. While both modalities are easy to access and feasible to use for a comprehensive muscle status in the acute setting of an ICU therapy, they are unable quantify muscle status in terms of fatty infiltration and muscle architecture in the long-term evaluation of ICU survivors [2, 15, 20]. Diagnosing ICU related polyneuropathy is standardized with electrophysiologic testing (EMG). However, ongoing long term muscle degeneration and myoarchitectural changes cannot be captured with it. Diagnostic MRI is the current gold-standard to evaluate the extend of acute and chronic muscle damage in neuropathies and myopathies and quantitative MR imaging (qMRI) protocols are increasingly used in neuromuscular research [21, 22]. qMRI markers such as mDixon fat-fraction, muscle diffusion tensor imaging and water T2 are established objective outcome measures to grade muscle degeneration and correlate with clinical muscle function [23–25]. Using mDixon sequences fatty infiltration can be objectively quantified, T2 mapping allows to capture muscle oedema and inflammation via water T2 relaxation time and mDTI can reveal myofiber atrophy on a microstructural level [25-29]. The result pattern of qMRI outcome measures allows conclusions on pathomechanisms of muscle degeneration and its classification as acute or chronic. Thus, an increased water T2 time reflects active muscle degeneration due to tissue edema, a fatty muscle infiltration can be measured with mDixon and reflects chronic muscle damage, and muscle fiber atrophy can be captured with the mDTI sequences [25-27]. Until now, there are no studies on long-term ICU survivors with or without ICUAW regarding qMRI evaluation. The aim of this study was to comprehensively evaluate the quality and quantity of structural muscle differences in long-term ICU survivors with a qMRI protocol and to correlate them to clinical findings and quality of life questionnaires. Better understanding of the long-lasting myoarchitectural abnormalities may contribute to optimized treatment options in the rehabilitation phase.

Methods

Study population

The subjects in the present study were part of a larger study (CRIT-Path study) with the aim to investigate the incidence of clinical and electrophysiological abnormalities in long-term ICU survivors. Subjects evaluated in the present study additionally underwent qMRI. Based on a medical patient management database screening, patients admitted to ICU between 2007 and 2017, (>7d on ICU of an university hospital including \geq 3d of invasive ventilation, at least 6mo-10y post-ICU, aged \leq 85y) were contacted by letter [3]. Volunteers then called the study centre to authorise study participation (see Table 1 for details). Sex- and age-matched controls without history of ICU treatment, neuromuscular diseases (NMD) or injuries in lower extremity 12 months before study enrolment were recruited via advertisement. MRI exclusion criteria were metal implants in lower legs or back or electronical implants such as a cochlear implant or drug pumps. This prospective study had been approved by the local ethics committee of the Ruhr-University Bochum (No. 4905-14 3.0) and written informed consent for participation and publication was obtained from all participants prior to enrolment.

Clinical and electrophysiological assessments

Muscle strength was evaluated using the Medical Research Council (MRC; 0–60) by an experienced clinician. Grip strength was measured with a hand

Name	Age (years)	Height (cm)	Weight (kg)	BMI	ICU stay (days)	Duration of ventilation (h)	distance (ICU to examination in days)	Anamnestic co-morbidities at study examination	
CP 1	77	187	94	26.88	10	95	1240	Hypertension, Coronary artery disease, obstructive sleep apnea	
CP 2	30	190	87	24.10	15	270	771	none	
CP 3	60	168	81	28.70	83	1848	712	Hypertension, Coronary artery disease	
CP 4	64	160	129	50.39	15	209	2422	Type-II-diabetes. Hyper-tension. Hypo- thyreoidism	
CP 5	54	178	138	43.56	19	369	965	Hypertension, Atrial fibrillation. Hypothy oidism	
CP 6	41	179	67	20.91	30	586	90	Hypertension. Coronary artery disease	
CP 7	47	186	83	23.99	42	240	6022	none	
CP 8	73	167	70	25.10	8	127	3588	Hypertension, Hyperthyroidism, Coronary artery disease	
CP 9	77	168	81	28.70	18	180	2555	Hypertension, Hypothyreoidism	

Table 1 Demographic data

ICU data, comorbidities; patients are labelled as CP 1-9

dynamometer on both hands [30]. Quality of life was measured with the EuroQOL5-Dimension questionnaire (EQ-5D-3L VAS). Subjects were assessed for any typical ICUAW symptoms like symmetrical proximal muscle weakness, distal dysesthesia, paraesthesia or pain (see Table 2 for details).

Electroneurography (ENG) was done unilaterally for the sural, the peroneal, the tibial and the ulnar nerve. For this study, we report tibial nerve compound motor action potential (TNCMAP) and sural nerve sensory action potential (SNSNAP). Polyneuropathy was graded as either axonal, demyelinating or combined based on the definition by England et al. [31] Electromyography (EMG) was performed unilaterally in the anterior tibialis

	Tab	le 2	Clinical	data
--	-----	------	----------	------

and the vastus lateralis muscle and graded as abnormal in
terms of acute, subacute or chronic neurogenic damage
according to Mills [32].

MRI acquisition and sequences

MR scans of both legs vertical to the femur and tibia bone were obtained using a Philips 3.0T Achieve MR system and a 16CH Torso XL coil. The participants lay in a feet-first supine position. Cushions were used to support participants' knees and sandbags placed around their feet to prevent motion.

For the first MRI acquisition protocol (first four patients) the thigh region from hip to knee was split into three fields of view (FOV) along the z-axis with a 30 mm

Name	ICUAW symptoms (yes/no)	Grip strength right (kg)	Grip strength left (kg)	MRC score	TNCMAP (mV)	SNSNAP (μV)	Type of polyneuropathy	EQ-5D-3L VAS
CP 1	Y	23	9	54	2.2	0	Axonal	50
CP 2	Ν	54	56	59	2.8	1.37	Mixed axonal demyelinating	90
CP 3	Υ	23	29	50	4.7	6.7	Mixed axonal demyelinating	50
CP 4	Ν	20	23	60	0	0	Axonal	100
CP 5	Y	24	21	56	3	4.2	Mixed axonal demyelinating	30
CP 6	Υ	11	12	52	7.1	8.7	Demyelinating	35
CP 7	Υ	39	26	58	8.8	15.3	Small fiber neuropathy	75
CP 8	Υ	28	30	60	8.5	9.5	Axonal in electromyography	90
CP 9	Y	32	28	59	2.1	0.83	Mixed axonal demyelinating	50

Grip strength was measured with a hand dynamometer in kilograms

MRC, Strength grade sum score according to Muscle Research Council: 0–60; TNCMAP, Compound muscle action potential of tibial nerve; SNSNAP, compound sensory nerve action potential of sural nerve; EQ-5D-3L VAS, EuroQoL 5-dimensional quality of life visual analogue scale ranging from 1 to 100

overlap, which each FOV comprised T1-weighted (T1w), T2-weighted (T2w), a diffusion-weighted spin-echo EPI (voxel size $3.0 \times 3.0 \times 6.0 \text{ mm}^3$; TR/TE 5000/57 ms; SPAIR/SPIR fat suppression; SENSE: 1.9; 17 gradient directions with b-values of 400 and 3 images with b-value of 0 [33] as well as one noise measurement (by turning of the RF and imaging gradients) with a total acquisition time of approximately 27 min for both thighs (9 min per FOV). An additional mDixonquant sequence (voxel size $1.5 \times 1.5 \times 6.0$ mm³; TR/TE 210/2.6, 3.36, 4.12, 4.88 ms; flip angle 8°, SENSE: 2) was acquired. After the image acquisition of the thigh-regions the data acquisition was paused and the TorsoXL coil was wrapped around the lower leg region; the calf region was split into two fields of view for additional 18 min scanning time for both calves.

The protocol for the remaining five patients consisted of a 4-point Dixon sequence (voxel size $1.5 \times 1.5 \times 6.0$ mm³; TR/TE 210/2.6, 3.36, 4.12, 4.88 ms; flip angle 8°, SENSE: 2), a multi-echo spin-echo (MESE) sequence for quantitative water mapping including 17 echoes and Cartesian k-space sampling (voxel size $3.0 \times 3.0 \times 6.0$ mm³; TR/TE 4598/17x Δ 7.6 ms; flip angle 90/180°, SENSE: 2), and a diffusion-weighted spin-echo EPI (voxel size $3.0 \times 3.0 \times 6.0$ mm³; TR/TE 5000/57 ms; SPAIR/SPIR fat suppression; SENSE: 1.9; 42 gradient directions with eight different b-values (0–600) [33]. A noise scan was obtained as described above. Here both, the thigh and the calf regions were both split into two fields of view each and the scanning time per stack was approximately 12 min.

Data pre-processing

Data from the first protocol were analysed as described previously in Schlaffke et al. [34, 35] Data from the second protocol were pre-processed as described before using QMRITools (www.qmritools.com) [33] In brief, the diffusion data were denoised using a PCA method [36]. To correct for subject motion and eddy currents both legs were registered separately. Then the tensors were calculated by taking IVIM into account and using an iWLLS algorithm. A non-linear IVIM fit of the diffusion data was performed as described in Orton et al. [37]. Furthermore, the IVIM bias signal was removed from diffusion weighted data using all acquired b-values [38]. By using IVIM correction, an isotropic pseudo-diffusion component was modelled in addition to the diffusion tensor, to effectively remove biases in mean diffusivity (MD) estimation.

However, if the pseudo-diffusion process was anisotropic and aligned with the orientation of the muscle fibers, this would result in an increase in fractional anisotropy (FA) independently from the IVIM correction [33, 39, 40]. The IDEAL method was used for the Dixon data considering a singleT2* decay and resulting in a separated water and fat map [41]. The derived water maps were used for the manual segmentation. Considering different T2 relaxation times for the water and fat components the T2-mapping data were processed using an extended phase graph (EPG) dictionary matching pattern method. Both water-T2 relaxation time and transmit B1 (B1+) were fitted for each voxel using a dictionary method. The T2 of fat Rwas obtained according to Marty et al. [42].

Muscle segmentation

Eight thigh muscles (vastus lateralis, vastus medialis, rectus femoris, semimembranosus, semitendinosus, biceps femoris, sartorius, and gracilis) and seven calf muscles (gastrocnemius medialis and lateralis, soleus, tibialis anterior, peroneus, extensor digitorum and tibialis posterior) were segmented manually avoiding subcutaneous fat and fascia on all slices of the reconstructed Dixon water images (3D-slicer 4.4.0, https://www.Slicer.org) [43].

The segmentations were then registered to T2 and DTI data to correct for small motions between sequences and image distortions using sequential rigid and b-spline transformations (elastix, https://elastix.lumc.nl) [44]. Average values within a muscle mask of water-T2 time (when available) and proton density fat fraction (FF) as well as the diffusion measures fractional anisotropy (FA), mean diffusivity (MD), radial diffusivity (RD) and axial diffusivity (λ_1) were obtained. SNR was calculated as the local average signal divided by the local noise sigma [45].

Outliers

Due to motion artifacts, image inhomogeneities and two patients aborted scanning, thigh data were obtained for 4/9 patients. Calf MRI data could be obtained in all included patients.

Statistical analysis

Water T2, FF, FA, MD, RD and λ_1 were compared between CIP patients and matched controls in a general linear model with patient/control as fixed factors as well as the protocol as nuisance variable, for all leg muscles.

To evaluate correlations between clinical assessments and qMRI values mean water T2, FF, FA, MD, RD and λ_1 of all thigh and calf muscles were correlated to grip strength, MRC, TNCMAP, SNSNAP, EQ-5D-3L VAS as well as level of symptoms using Pearson's correlation coefficients. All statistical analyses were performed using IBM SPSS V28. The significance level for all tests was set at p < 0.05.

Results

Patent cohort

In total, 9 long-term ICU survivors (7 males; 54.8 ± 16.9 y) as well as age and gender matched controls (7 males; $54.3y \pm 11.14y$) were included. Mean duration of ICU therapy was 26.2 ± 22.2 days. The mean interval between ICU therapy and testing was 5.4 ± 4.8 y. Further demographic data, ICU data and comorbidities of patients can be found in Table 1. Clinical data including persisting symptoms, data on quality of life (EQ-5D-3L), grip-strength, muscle strength by MRC sum score, electrophysiological data (TNCMAP, SNSNAP) and polyneuropathy classification are listed in Table 2. In all included ICU-survivors a polyneuropathy was diagnosed.

qMRI outcome measures in ICU survivors compared to controls

Water T2 mapping sequence was acquired for 5/9 patients due to a change in examination protocol (see methods). Example images of the applied MRI sequences are shown in Fig. 1. The average qMRI values were significantly higher in all patient muscles compared to controls (multivariate general linear model: main effect: p < 0.001 for water T2, FF, MD, λ_1 , RD;

p-value for FA was 0.047; see Table 3). The mean qMRI values of all muscles combined for patients and controls are displayed in Fig. 2. For complete qMRI data see Table S1.

When only thigh muscles were compared between ICU survivors and controls only FF, MD, λ_1 and RD were significantly higher and FA and water T2 were not significant (multivariate general linear model with examination protocol as covariate—main effect: p < 0.001 for FF and λ_1 ; for FA: p = 0.357; for MD: p = 0.001; for λ_2 : p = 0.002, for λ_3 : p = 0.038, for water T2: p = 0.957, for RD: p = 0.009; See also Table 3). The mean qMRI values (FA, MD, FF, water T2, RD) of thigh muscles combined for patients and controls are displayed in Fig. 3.

When calf muscles were compared between ICU survivors and controls water T2, FF, MD, λ_1 , RD were significantly higher (multivariate general linear model with examination protocol as covariate—main effect: p < 0.001 for MD, λ_1 , water T2, RD; for FF: p = 0.022; for FA: p = 0.429; see also Table 3). The mean qMRI values of calf muscles combined for patients and controls are displayed in Fig. 4.



Fig. 1 Example images of the applied MRI sequences: mDixon fat fraction (FF), fractional anisotropy (FA), mean diffusivity (MD) and water T2 maps for thigh and calf muscles of two representative patients, and a healthy control (HC)

Table 3 Multivariate general linear model results of qMRI parameters between controls (CON) and patients (CIP)

q-MRI parameters	All musc	les	Thigh muscles	Calf muscles	
n muscles (PAT/ CON)	98/300		28/160	70/140	
T2	∆mean	2.6	0.01	3.33	
ms	p value	< 0.001*	0.957	<0.001*	
n muscles (PAT/ CON)	164/300		52/160	112/140	
FF	∆mean	0.79	0.71	2.2	
%	p value	<0.001*	< 0.001*	0.022*	
FA	∆mean	0.02	0.025	0.01	
	p value	0.047*	0.352	0.429	
MD	∆mean	0.1	0.06	0.11	
$[10^{-3} \text{ mm}^2/\text{s}]$	p value	<0.001*	0.001*	<0.001*	
λ ₁	∆mean	0.16	0.12	0.15	
$[10^{-3} \text{ mm}^2/\text{s}]$	p value	< 0.001*	< 0.001*	<0.001*	
RD	∆mean	0.07	0.03	0.1	
$[10^{-3} \text{ mm}^2/\text{s}]$	p value	<0.001*	0.009*	<0.001*	

Significant results are highlighted with * and expressed in bold. Significance level was calculated as p < 0.05. Δ -mean was calculated as: Mean of PAT value–Mean of CON value, for each parameter

FF, fat fraction; MD, mean diffusivity; FA, fractional anisotropy; RD, radial diffusivity; T2, water T2-time

Water T2 was significantly higher in calf muscles of ICU survivors compared to controls (p < 0.001). In contrast in thigh muscles there were no differences

between ICU survivors and controls (p = 0.957; See Table 3, Figs. 2, 3 and 4).

Detailed qMRI values for each muscle separately can be found in Additional file 1: Table S1.

Correlation between qMRI and clinical data

Correlations between clinical assessments and qMRI values in patients are displayed in Table 4. A significant negative correlation between MRC and FA (r = -0.75, p = 0.02) as well as between MRC and water T2 (r = -0.986, p = 0.002) could be observed for all muscles (see Fig. 5 for correlation plots). Furthermore, a significant negative correlation between water T2 and grip strength with significance for the left hand (r = -0.987, p = 0.002) was revealed. Especially water T2 showed a strong negative linear correlation with MRC grade. FF showed a negative correlation with tibial nerve compound motor action potential (r = -0.719, p = 0.029). No significant correlations could be found between other qMRI values and clinical as well as electrophysiological or quality of life assessments.

Discussion

In this pilot study we show that qMRI values in leg muscles of patients after ICU treatment differ significantly from controls even years after ICU therapy and could reflect simultaneously muscle damage and chronic myostructural abnormalities [27–29, 46]. Water T2 time and FA correlate negatively with MRC sum score, indicating the clinical relevance of our findings.



Fig. 2 Box-plot of qMRI values of FA, MD, RD, T2 and Fat Fraction for all muscles displayed with a min to max range. See also Table 3 for p-values. FF, fat fraction; MD, mean diffusivity; FA, fractional anisotropy; RD, radial diffusivity; T2, water T2-time. Pat = ICU survivors; Con = Controls



Fig. 3 Box-plot of qMRI values of FA, MD, RD, T2 and Fat Fraction for all thigh muscles displayed with a min to max range. See also Table 3 for p-values. FF, fat fraction; MD, mean diffusivity; FA, fractional anisotropy; RD, radial diffusivity; T2, water T2-time. Pat = ICU survivors; Con = Controls



Fig. 4 Box-plot of qMRI values of FA, MD, RD, T2 and Fat Fraction for calf muscles displayed with a min to max range. See also Table 3 for p-values. FF, fat fraction; MD, mean diffusivity; FA, fractional anisotropy; RD, radial diffusivity; T2, water T2-time. Pat = ICU survivors; Con = Controls

An elevated FF, as observed in our cohort, relates to chronic muscle degeneration and shows higher sensitivity compared to MRC testing and qualitative MRI in terms of capturing muscle degeneration in myopathies [26, 47]. Fatty infiltration is a sign of previous muscle damage with fatty replacement of irreversible damaged muscle fibers [48]. A high FF in muscle supports the hypothesis that long-term motor dysfunction and muscle fatigue in patients is caused by a myostructural deficit [2, 3, 14, 49]. FF values were only slightly elevated in ICU survivors compared to controls and still in a "normal" range. Whereas FF describes fatty infiltration very accurately and is easy to capture as percent fat per muscle, changes in the other qMRI modalities DTI and water

qMRI	Grip strength right	Grip strength left	MRC	TNCMAP	SNSNAP	EQ5DVAS	Pain	Paresthesia
FF								
Pearson	276	130	.148	719*	553	.100	043	.120
p-value	.472	.739	.703	.029*	.123	.799	.913	.759
n	9	9	9	9	9	9	9	9
MD								
Pearson	008	— .337	.204	450	284	309	131	.078
p-value	.983	.375	.599	.224	.458	.419	.738	.841
n	9	9	9	9	9	9	9	9
FA								
Pearson	— .510	— .132	750*	501	515	— .610	491	165
p-value	.161	.734	.020*	.169	.156	.081	.180	.672
n	9	9	9	9	9	9	9	9
RD								
Pearson	.133	283	.288	329	— .178	314	123	.031
p-value	.733	.460	.453	.387	.647	.410	.752	.938
n	9	9	9	9	9	9	9	9
T2								
Pearson	862	987**	986**	.088	.064	648	.142	.486
p-value	.060	.002*	.002*	.888	.918	.237	.820	.406
n	5	5	5	5	5	5	5	5

Table 4 Correlations of qMRI parameters with clinical parameters

Pearson correlation level; p-value and number of subjects (n) are presented. Significance level was set at p < 0.05. Significant values are presented in bold and highlighted with *. MRC = Muscle Research Council strength grade (0–60)

TNCMAP, tibial nerve compound action potential; SNSSNAP, Sensory neve compound action potential; FF, fat fraction; MD, mean diffusivity; FA, fractional anisotropy; RD, radial diffusivity; T2, water T2-time



Fig. 5 Correlation plots muscle strength by MRC (0–60) versus FA and MRC versus T2. FA, fractional anisotropy, FF, fat fraction; MRC, Medical Research Council

T2 need to be closely interpreted with the expected or known disease pathology.

Whereas CIM is transient and only reflects myostructural damage due to direct muscle injury in the acute phase of ICU therapy, CIP in contrast is long-lasting due to nerve injury and serves as the major cause of functional debilitation [50]. Neurogenic myofiber atrophy due to axonal loss is usually irreversible and has already been described in muscle biopsies of CIP patients [51]. Our electrophysiologic studies show predominantly axonal nerve damage in calf nerves of patients and clinical data underline that CIP usually affects distal limbs [51]. Since we confirmed a polyneuropathy in our patient cohort, consistent with CIP, our observed differences of qMRI parameters reflect neurogenic muscle damage. The observed higher FA in patient muscles shows a higher proportion of axial compared to radial diffusion. In neurogenic myofiber atrophy,

myofibers do not lose their structural integrity initially but get atrophic. This atrophy leads to myofiber diameter reduction and consequently to an increase in FA [29, 52]. MD usually reflects the degree of overall diffusion as it equally integrates the eigenvalues $\lambda_1 - \lambda_3$ as a simple mean value (MD = $\frac{\sum(\lambda_1,\lambda_2,\lambda_3)}{3}$) and is elevated in active muscle degeneration and inflammation whereas a reduction of MD is seen in muscle atrophy without an active degeneration. The relation of MD and FA values is usually fixed [29]. Thus a solely myofiber atrophy would lead to an increase in FA and a decrease in MD whereas an inflammatory edema would counterwise lead to a decrease in FA and an increase in MD (as well as RD, and $\lambda_{2\text{--}3})$ [29]. Interestingly, in our patients, FA and MD are both significantly higher than in controls mostly pronounced in the calf muscles. A combination of a high FA and a high MD supports a parallel chronic myofiber atrophy and active myofiber degeneration.

Underlying active muscle fiber degeneration are revealed by a significantly elevated water T2 which is usually elevated in tissue with a higher content of fluid (e.g. inflammation, myofiber breakdown) [53–55].

Thus, we hypothesize that our observed combination of unanimous elevated MD, water T2, FA and FF reflect parallel active muscle degeneration and myofiber atrophy in chronic damaged muscle tissue due to ICUAW. Although FF is significantly increased between the two groups, the absolute value is still comparably low, so that an influence of the FF on the DTI parameters can be ruled out [56, 57]. This hypothesis is also underlined by a separate analysis of thigh and calf muscle qMRI values and by our clinical outcomes.

In thighs, compared to calves, chronic muscle degeneration is predominantly observed as MD is only mildly elevated compared to calves and water T2 is not elevated. In contrast, MD and water T2 are highly elevated in calves and reflect active muscle degeneration. Our data support that chronic axonal nerve damage in calves due to CIP leads to an ongoing myofiber damage and breakdown.

Correlations of FA and MD to clinical assessments have been described in myopathies before and the significant correlation of water T2 and FA to MRC values in our study additionally supports the relevance of qMRI values [57, 58]. Since up to now there are no qMRI studies in long-term ICU survivors the presented results and the discussed pathophysiology is derived from known relations between quantitative muscle MRI parameters and myofiber degeneration. Reviews on ICUAW and known long-term data on ICU-survivors highlight the impact of motor status on long term quality of life [59], since ICUAW and especially CIP affects peripheral nerves irreversibly and leads to ongoing neuromuscular complaints and a reduced ability to participate in daily life activities.

Conclusion

We conclude that using qMRI we were able to quantify clinically relevant muscle differences in ICU survivors probably due to an ICUAW. These findings can help to characterize underlying mechanisms for ongoing neuromuscular complaints in long-term ICU-survivors. qMRI parameters show chronic next to active muscle degeneration in ICU survivors with diagnosed CIP according to the electrophysiological assessment within the study.

Limitations

Since our recruitment was narrow and patients with long-term ICU data are not easy to recruit and are frequently affected with confounding diseases or metal implants most potential subjects did not meet our inclusion criteria. The acquisition protocols was changed during the study for protocol optimization and the implementation of T2-mapping. The protocol was integrated as a covariate in statistical analysis to correctly minimize the statistical impact on the results. Due to outliers in data acquisition only four thighs were available for statistical analysis. This may have confounded significance levels and differences might be induced in diffusion values between calves and thighs. FA was not elevated when thigh and calf muscles were analysed separately. Those different findings compared to the analysis of all muscles may be explained by methodological reasons and the small number of subjects.

Abbreviations

CIM	Critical illness myopathy
CIP	Critical illness polyneuropathy
DTI	Diffusion tensor imaging
EQ-5D-3L	EuroQOL5-Dimension questionnaire
FA	Fractional anisotropy
FF	Fat fraction
FOV	Field of view
ICU	Intensive care unit
ICUAW	Intensive care unit acquired weakness
MD	Mean diffusivity
MRC	Medical Research Council
MRI	Magnetic resonance imaging
NMD	Neuromuscular diseases
qMRI	Quantitative magnetic resonance imaging
RD	Radial diffusivity
SNR	Signal to noise ratio
SNSNAP	Sural nerve action potential
T2	Water T2
TNCMAP	Tibial nerve compound motor action potential
VAS	Visual analogue scale

Supplementary Information

The online version contains supplementary material available at https://doi. org/10.1186/s12880-023-00995-7.

Additional file 1. Supplementary Table.

Acknowledgements

The authors thank EuroQOL Group for written permission to use the EQ-5D-3L. We thank Philips Germany especially Burkhard Mädler for continuous scientific support and Melanie Ebel for her comprehensive support.

Author contributions

RR was involved in data acquisition, statistics, patient selection, clinical assessment, manuscript preparation and finalization, creation of figures and tables as well as statistical analysis; LS was involved in study conceptualization, data acquisition, statistics, manuscript finalization as well as statistical analysis; EK was involved in data research, statistics, patient selection, manuscript preparation and finalization and clinical assessment. CMF was involved in study conceptualization, data acquisition, statistics, patient selection, manuscript preparation and finalization and funding. All authors read and approved the final manuscript.

Funding

Open Access funding enabled and organized by Projekt DEAL. This research work was supported by means of the German Social Accident Insurance (DGUV) and FoRUM of the Ruhr University Bochum, Germany. EEK holds an endowed professorship funded by the German Social Accident Insurance (DGUV) for the period of 6 years (2020–2020) and has received a grant from the Georg Agricola Ruhr foundation. LS receives funding from the Deutsche Forschungsgemeinschaft (DFG—SFB 874: Project: TP-A5, No. 122679504). The funding bodies played no role in the design of the study and collection, analysis, interpretation of data, and in writing the manuscript. We acknowledge support by the Open Access Publication Funds of the Ruhr-Universität Bochum.

Availability of data and materials

The data that support the findings of this study are not openly available due to sensitivity of human data and to protect patient privacy. The data are available from the corresponding author upon written reasonable request. Any written request will be reviewed by the data protection officer of the University Hospital Bergmannsheil Bochum prior to access.

Declarations

Ethics approval and consent to participate

This prospective study had been approved by the local ethics committee of the Ruhr-University Bochum (No. 4905-14 3.0). All methods were carried out in accordance with Declaration of Helsinki. Written informed consent for participation was obtained from all participants prior to enrolment.

Consent for publication

Not applicable.

Competing interests The authors declare that they have no competing interests.

Author details

¹Department of Neurology, BG-University Hospital Bergmannsheil gGmbH, Ruhr-University Bochum, Bürkle-de-La-Camp-Platz 1, 44789 Bochum, Germany. ²Department of Anaesthesiology, Intensive Care and Pain Medicine, BG-University Hospital Bergmannsheil, Ruhr-University Bochum, Bochum, Germany.

Received: 20 September 2022 Accepted: 8 March 2023 Published online: 18 March 2023

References

- Hermans G, Van den Berghe G. Clinical review: intensive care unit acquired weakness. Crit Care. 2015;19(1):1–9.
- 2. Kress JP, Hall JB. ICU-acquired weakness and recovery from critical illness. N Engl J Med. 2014;370(17):1626–35.
- Meyer-Frießem CH, Malewicz NM, Rath S, Ebel M, Kaisler M, Tegenthoff M, et al. Incidence, time course and influence on quality of life of intensive care unit-acquired weakness symptoms in long-term intensive care survivors. J Intensive Care Med. 2021;36(11):1313–22.
- Derde S, Hermans G, Derese I, Güiza F, Hedström Y, Wouters PJ, et al. Muscle atrophy and preferential loss of myosin in prolonged critically ill patients. Crit Care Med. 2012;40(1):79–89.
- Bloch SA, Lee JY, Syburra T, Rosendahl U, Griffiths MJD, Kemp PR, et al. Increased expression of GDF-15 may mediate ICU-acquired weakness by down-regulating muscle microRNAs. Thorax. 2015;70(3):219–28.
- Corpeno Kalamgi R, Salah H, Gastaldello S, Martinez-Redondo V, Ruas JL, Fury W, et al. Mechano-signalling pathways in an experimental intensive critical illness myopathy model. J Physiol. 2016;594(15):4371–88.
- Fletcher SN, Kennedy DD, Ghosh IR, Misra VP, Kiff K, Coakley JH, et al. Persistent neuromuscular and neurophysiologic abnormalities in long-term survivors of prolonged critical illness. Crit Care Med. 2003;31(4):1012–6.
- Herridge MS, Tansey CM, Matté A, Tomlinson G, Diaz-Granados N, Cooper A, et al. Functional disability 5 years after acute respiratory distress syndrome. N Engl J Med. 2011;364(14):1293–304.
- Adhikari NK, Fowler RA, Bhagwanjee S, Rubenfeld GD. Critical care and the global burden of critical illness in adults. The Lancet. 2010;376(9749):1339–46.
- McNelly AS, Rawal J, Shrikrishna D, Hopkinson NS, Moxham J, Harridge SD, et al. An exploratory study of long-term outcome measures in critical illness survivors: construct validity of physical activity, frailty, and healthrelated quality of life measures. Crit Care Med. 2016;44(6):e362-369.
- Batt J, dos Santos CC, Cameron JI, Herridge MS. Intensive care unitacquired weakness. Am J Respir Crit Care Med. 2013;187(3):238–46.
- Dos Santos C, Hussain SNA, Mathur S, Picard M, Herridge M, Correa J, et al. Mechanisms of chronic muscle wasting and dysfunction after an intensive care unit stay. A pilot study. Am J Respir Crit Care Med. 2016;194(7):821–30.
- Hermans G, Van Mechelen H, Bruyninckx F, Vanhullebusch T, Clerckx B, Meersseman P, et al. Predictive value for weakness and 1-year mortality of screening electrophysiology tests in the ICU. Intensive Care Med. 2015;41(12):2138–48.
- Van Aerde N, Meersseman P, Debaveye Y, Wilmer A, Gunst J, Casaer MP, et al. Five-year impact of ICU-acquired neuromuscular complications: a prospective, observational study. Intensive Care Med. 2020;46(6):1184–93.
- Puthucheary ZA, Phadke R, Rawal J, McPhail MJW, Sidhu PS, Rowlerson A, et al. Qualitative ultrasound in acute critical illness muscle wasting. Critical Care Med. 2015;43(8):1603–11.
- Seymour JM, Ward K, Sidhu PS, Puthucheary Z, Steier J, Jolley CJ, et al. Ultrasound measurement of rectus femoris cross-sectional area and the relationship with quadriceps strength in COPD. Thorax. 2009;64(5):418–23.
- Yildiz OK, Yildiz B, Avci O, Hasbek M, Kanat S. Clinical, neurophysiological and neuroimaging findings of critical illness myopathy after COVID-19. Cureus. 2021;13(3). Verfügbar unter: https://www.cureus.com/articles/ 53380-clinical-neurophysiological-and-neuroimaging-findings-of-criticalillness-myopathy-after-covid-19
- Fisse AL, May C, Motte J, Pedreiturria X, Breuer TGK, Schneider-Gold C, et al. New approaches to critical illness polyneuromyopathy: high-resolution neuromuscular ultrasound characteristics and cytokine profiling. Neurocrit Care. 2021;35(1):139–52.
- Paolo F, Valentina DG, Silvia C, Tommaso P, Elena C, Martin D, et al. The possible predictive value of muscle ultrasound in the diagnosis of ICUAW in long-term critically ill patients. J Crit Care. 2022;71:154104.
- Grimm A, Teschner U, Porzelius C, Ludewig K, Zielske J, Witte OW, et al. Muscle ultrasound for early assessment of critical illness neuromyopathy in severe sepsis. Crit Care. 2013;17(5):R227.
- Tomas X, Milisenda JC, Garcia-Diez AI, Prieto-Gonzalez S, Faruch M, Pomes J, et al. Whole-body MRI and pathological findings in adult patients with myopathies. Skelet Radiol. 2019;48(5):653–76.

- Caetano AP, Alves P. Advanced MRI patterns of muscle disease in inherited and acquired myopathies: what the radiologist should know. Semin Musculoskelet Radiol. 2019;23(03):e82-106.
- Paoletti M, Pichiecchio A, Cotti Piccinelli S, Tasca G, Berardinelli AL, Padovani A, et al. Advances in quantitative imaging of genetic and acquired myopathies: clinical applications and perspectives. Front Neurol. 2019;10. Verfügbar unter: https://www.ncbi.nlm.nih.gov/pmc/articles/PMC63 78279/
- Klickovic U, Zampedri L, Sinclair CDJ, Wastling SJ, Trimmel K, Howard RS, et al. Skeletal muscle MRI differentiates SBMA and ALS and correlates with disease severity. Neurology. 2019;93(9):e895–907.
- Damon BM, Li K, Bryant ND. Magnetic resonance imaging of skeletal muscle disease. Handb Clin Neurol. 2016;136:827–42.
- Arrigoni F, De Luca A, Velardo D, Magri F, Gandossini S, Russo A, et al. Multiparametric quantitative MRI assessment of thigh muscles in limbgirdle muscular dystrophy 2A and 2B: quantitative MRI in LGMD2A and LGMD2B. Muscle Nerve. 2018;58(4):550–8.
- Güttsches AK, Rehmann R, Schreiner A, Rohm M, Forsting J, Froeling M, et al. Quantitative muscle-MRI correlates with histopathology in skeletal muscle biopsies. J Neuromuscul Dis. 2021;8(4):669–78.
- Rehmann P, Schneider-Gold C, Froeling M, Güttsches AK, Rohm M, Forsting J, et al. Diffusion tensor imaging shows differences between myotonic dystrophy type 1 and type 2. J Neuromuscul Dis. 2021;8(6):949–62.
- Berry DB, Regner B, Galinsky V, Ward SR, Frank LR. Relationships between tissue microstructure and the diffusion tensor in simulated skeletal muscle. Magn Reson Med. 2017.
- Hermans G, Clerckx B, Vanhullebusch T, Segers J, Vanpee G, Robbeets C, et al. Interobserver agreement of medical research council sumscore and handgrip strength in the intensive care unit. Muscle Nerve. 2012;45(1):18–25.
- England JD, Gronseth GS, Franklin G, Miller RG, Asbury AK, Carter GT, et al. Distal symmetrical polyneuropathy: definition for clinical research. Muscle Nerve. 2005;31(1):113–23.
- Mills KR. The basics of electromyography. J Neurol Neurosurg Psychiatry. 2005;76(suppl 2):ii32–5.
- Schlaffke L, Rehmann R, Rohm M, Otto LAM, De Luca A, Burakiewicz J, et al. Multi-center evaluation of stability and reproducibility of quantitative MRI measures in healthy calf muscles. NMR Biomed. 2019;32(9):e4119.
- Rehmann R, Froeling M, Rohm M, Forsting J, Kley RA, Schmidt-Wilcke T, et al. Diffusion tensor imaging reveals changes in non-fat infiltrated muscles in late onset Pompe disease. Muscle Nerve. 2020.
- Rehmann R, Schlaffke L, Froeling M, Kley RA, Kühnle E, De Marées M, et al. Muscle diffusion tensor imaging in glycogen storage disease V (McArdle disease). Eur Radiol. 2019;29(6):3224–32.
- Leemans A, Jones DK. The B-matrix must be rotated when correcting for subject motion in DTI data. Magn Reson Med. 2009;61(6):1336–49.
- Orton MR, Collins DJ, Koh DM, Leach MO. Improved intravoxel incoherent motion analysis of diffusion weighted imaging by data driven Bayesian modeling: improved IVIM analysis with Bayesian modelling. Magn Reson Med. 2014;71(1):411–20.
- De Luca A, Bertoldo A, Froeling M. Effects of perfusion on DTI and DKI estimates in the skeletal muscle. Magn Reson Med. 2016.
- Veraart J, Novikov DS, Christiaens D, Ades-aron B, Sijbers J, Fieremans E. Denoising of diffusion MRI using random matrix theory. Neuroimage. 2016;142:394–406.
- Veraart J, Sijbers J, Sunaert S, Leemans A, Jeurissen B. Weighted linear least squares estimation of diffusion MRI parameters: strengths, limitations, and pitfalls. Neuroimage. 2013;81:335–46.
- Reeder SB, Pineda AR, Wen Z, Shimakawa A, Yu H, Brittain JH, et al. Iterative decomposition of water and fat with echo asymmetry and least-squares estimation (IDEAL): application with fast spin-echo imaging. Magn Reson Med. 2005;644:636–44.
- Marty B, Baudin PY, Reyngoudt H, Azzabou N, Araujo ECAA, Carlier PG, et al. Simultaneous muscle water T2 and fat fraction mapping using transverse relaxometry with stimulated echo compensation. NMR Biomed. 2016;29:431–43.
- Rehmann R, Schlaffke L, Froeling M, Kley RA, Kühnle E, De Mareés M, et al. Muscle diffusion tensor imaging in Glycogen storage disease V (McArdle disease). Eur Radiol. 2018;29(6):3224–32.

- Klein S, Staring M, Murphy K, Viergever MA, Pluim J. elastix: A toolbox for intensity-based medical image registration. IEEE Trans Med Imaging. 2010;29(1):196–205.
- Froeling M, Tax CMW, Vos SB, Luijten PR, Leemans A. "MASSIVE" brain dataset: multiple acquisitions for standardization of structural imaging validation and evaluation. Magn Reson Med. 2017;77(5):1797–809.
- Oudeman J, Nederveen AJ, Strijkers GJ, Maas M, Luijten PR, Froeling M. Techniques and applications of skeletal muscle diffusion tensor imaging: a review. J Magn Reson Imaging. 2015;43:773–88.
- Carlier PG, Marty B, Scheidegger O, De Sousa PL, Baudin PY, Snezhko E, et al. Skeletal muscle quantitative nuclear magnetic resonance imaging and spectroscopy as an outcome measure for clinical trials. J Neuromusc Dis. 2016;3(1):1–28.
- Willis TA, Hollingsworth KG, Coombs A, Sveen ML, Andersen S, Stojkovic T, et al. Quantitative muscle MRI as an assessment tool for monitoring disease progression in LGMD2I: a multicentre longitudinal study. PLoS ONE. 2013;8(8):e70993.
- Needham DM, Davidson J, Cohen H, Hopkins RO, Weinert C, Wunsch H, et al. Improving long-term outcomes after discharge from intensive care unit: report from a stakeholders' conference. Critical Care Med. 2012;40(2):502–9.
- Zhou C, Wu L, Ni F, Ji W, Wu J, Zhang H. Critical illness polyneuropathy and myopathy: a systematic review. Neural Regen Res. 2014;9(1):101–10.
- Latronico N, Bolton CF. Critical illness polyneuropathy and myopathy: a major cause of muscle weakness and paralysis. Lancet Neurol. 2011;10(10):931–41.
- Otto LAM, Froeling M, Eijk RPA, Asselman F, Wadman R, Cuppen I, et al. Quantification of disease progression in spinal muscular atrophy with muscle MRI—a pilot study. NMR Biomed. 2021. https://doi.org/10.1002/ nbm.4473.
- Stouge A, Khan KS, Kristensen AG, Tankisi H, Schlaffke L, Froeling M, et al. MRI of Skeletal Muscles in Participants with Type 2 Diabetes with or without Diabetic Polyneuropathy. Radiology. 2020;297(3):608–19.
- Carlier PG, Azzabou N, de Sousa PL, Hicks A, Boisserie JM, Amadon A, et al. Skeletal muscle quantitative nuclear magnetic resonance imaging followup of adult Pompe patients. J Inherit Metab Dis. 2015;38(3):565–72.
- Damon BM, Froeling M, Buck AKW, Oudeman J, Ding Z, Nederveen AJ, et al. Skeletal muscle diffusion tensor-MRI fiber tracking: rationale, data acquisition and analysis methods, applications and future directions. NMR Biomed. 2016;30:e3563.
- Damon BM, Ding Z, Anderson AW, Freyer AS, Gore JC. Validation of diffusion tensor MRI-based muscle fiber tracking. Magn Reson Med. 2002;48(1):97–104.
- 57. Otto LAM, van der Pol WL, Schlaffke L, Wijngaarde CA, Stam M, Wadman RI, et al. Quantitative MRI of skeletal muscle in a cross-sectional cohort of patients with spinal muscular atrophy types 2 and 3. NMR in Biomed. 2020;33:e4357:1-e4413.
- Hooijmans MT, Damon BM, Froeling M, Versluis MJ, Burakiewicz J, Verschuuren JJGM, et al. Evaluation of skeletal muscle DTI in patients with duchenne muscular dystrophy. NMR Biomed. 2015;28:1589–97.
- Intiso D, Centra AM, Bartolo M, Gatta MT, Gravina M, Di Rienzo F. Recovery and long term functional outcome in people with critical illness polyneuropathy and myopathy: a scoping review. BMC Neurol. 2022;22(1):50.

Publisher's Note

Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.